# SOYASAPONIN A3, A NEW MONODESMOSIDIC SAPONIN ISOLATED FROM THE SEEDS OF GLYCINE MAX

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ABSTRACT.—A new triterpenoid saponin, named soyasaponin A3, has been isolated from soyabean and identified as 3-0-[ $\alpha$ -L-rhamnopyranosyl(1 $\rightarrow$ 2)- $\beta$ -D-galactopyranosyl(1 $\rightarrow$ 2)- $\beta$ -D-glucuronopyranosyl]olean-12-en-21 $\beta$ ,22 $\beta$ ,24-triol on the basis of chemical and spectral evidence.

The presence of saponins in foodstuffs has attracted considerable interest over the past few years (1,2) due to their physiological properties, including hypocholesterolemic activity (3), gut wall permeabilization (4), and bitter taste (5). During the course of our studies in this area, we have isolated and characterized three additional saponins from soyabean [*Glycine max* (L.) Merrill, Leguminosae] in addition to those previously reported [soyasaponins I, II, III, and A<sub>1</sub> and A<sub>2</sub>, which have been fully characterized by Kitagawa and co-workers (6–8)]. Two of these additional saponins, soyasaponins IV and V, the monodesmosides of soyasapogenol B, have been previously reported (9,10) while the third, tentatively named soyasaponin A<sub>3</sub> [1], a monodesmoside of soyasapogenol A, is described here.

## RESULTS

The saponin **1** was eluted from a Si gel column as a mixture with soyasaponins I and V as described previously (9). It was purified by further chromatography using a reversed-phase ( $KC_{18}$ ) flash chromatography column to give a product homogeneous on both normal and reversed-phase tlc.

Fabms of the intact saponin 1 in the negative mode gave a molecular ion at m/z 957  $[M-H]^-$  and ions at m/z 811 [M-H- deoxyhexose]<sup>-</sup>, 649 [M-H- deoxyhexose – hexose]<sup>-</sup>, and 473 [M-H- deoxyhexose – hexose – hexoronic acid]<sup>-</sup>. The spectrum



obtained in the positive mode gave a protonated molecular ion at m/z 959 together with a sodium adduct at m/z 981 and ions at m/z 813 [M+H- deoxyhexose]<sup>+</sup>, m/z 651 [M+H- deoxyhexose - hexose]<sup>+</sup>, and at m/z 457, 439, and 421, indicative of losses of one, two, and three molecules of H<sub>2</sub>O, respectively, from a saponin containing the aglycone, soyasapogenol A (mol wt 474). Gc of the trimethylsilyl derivative of the acid hydrolysis product of the saponin confirmed the presence of soyasapogenol A.

The elution characteristics of the saponin on normal phase tlc, where it coeluted with soyasaponin I, were indicative of a monodesmosidic structure. On reversed-phase tlc,  $\mathbf{1}$  ( $R_f$  0.35) eluted between soyasaponin I ( $R_f$  0.17) and the bisdesmosidic soyasaponin  $A_2$  ( $R_f$  0.48) showing an intermediate polarity consistent with the structure suggested.

Standard sugar analysis and gc of the resultant alditol acetates showed the presence of rhamnose and galactose in a ratio of 2:1. <sup>1</sup>H nmr of the permethylated saponin gave signals at  $\delta$  4.28 (d, J=7.5 Hz),  $\delta$  4.65 (d, J=7.8 Hz), and  $\delta$  5.24 (s) indicating the presence of  $\beta$ -linked glucuronic acid and galactose and an  $\alpha$ -linked rhamnose, respectively.

Coupled gc-ms of the partially methylated alditol acetates gave three components characteristic of a terminal rhamnose (Rt=79 min and m/z 175, 162, 131, 118), a 1,2 galactose (Rt=202 min and m/z 234, 205, 190, 161), and a deuterated 1,2-glucose (Rt=257 min and m/z 235, 191, 190). On the basis of these results the structure of the saponin was concluded to be 3-0-[ $\alpha$ -L-rhamnopyranosyl(1 $\mapsto$ 2)- $\beta$ -D-galactopyranosyl (1 $\mapsto$ 2)- $\beta$ -D-glucuronopyranosyl]olean-12-en-21 $\beta$ ,22 $\beta$ ,24-triol. The existence of **1** comprising the carbohydrate moiety of soyasaponin I and the aglycone soyasapogenol A common to both soyasaponins A<sub>1</sub> and A<sub>2</sub> is consistent with previously suggested biosynthetic pathways (11).

## EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Reversed-phase flash chromatography column packing was obtained from J.T. Baker Chemical Co. Ltd., Phillipsburg, New Jersey. Reversed phase tlc plates were supplied by Whatman Separation Ltd. Fabms were obtained using a KRATOS MS80 RFA instrument (having a potential of 5–8 kV applied to the xenon gun). The <sup>1</sup>H nmr (400 MHz) was obtained on a JEOL GX400 spectrometer in  $C_5D_5N$ . Gc of the aglycone-trimethylsilyl derivative was carried out on a Carlo Erba HRGC 5300 chromatograph with FID using a 3% OV-1 column at 270°. Gc separation of the alditol acetates was carried out on a Carlo Erba HRGC 5300 with FID, and combined gc-ms of the partially methylated alditol acetates was carried out on a Perkin-Elmer Sigma 2 instrument coupled to an MS30 mass spectrometer, both using a 3% OV-225 on Diatomite C2 (100–200 mesh, 1.5 m×2.3 mm i.d.) column isothermally at 210° or programmed between 150° and 220° at 1°/min. Tlc was performed using normal phase Si gel plates (*n*-BuOH-EtOH-NH<sub>3</sub>; 7:2:5), Si gel plates modified by oxalic acid (CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O; 65:35:10 lower layer), and reversed-phase KC<sub>18</sub> octadecylsilane bonded to Si gel (MeOH-H<sub>2</sub>O; 3:2).

EXTRACTION PROCEDURES.—Soyabean meal (100 g) was defatted by Soxhlet extraction with CHCl<sub>3</sub> (800 ml, 16 h). The air-dried, defatted meal was then Soxhlet-extracted with redistilled MeOH (900 ml, 30 h). The MeOH extract was concentrated to dryness, dissolved in  $H_2O$  (50 ml), and eluted through a column of octadecylsilane ( $C_{18}$ ) bonded Si gel (100 g) with  $H_2O$  (500 ml) and MeOH (500 ml). The MeOH eluate was evaporated to dryness (1 g) and treated with 5% methanolic KOH by reflux (30 min) to remove any acetyl groups. The reaction mixture was neutralized with ion exchange resin (Dowes 50W), filtered, evaporated to dryness, and applied to a Si gel column (100 g) in CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (7:3:1) and eluted with the same solvent. Fractions were monitored by tlc on Si gel (COOH)<sub>2</sub> (CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O; 65:35:10, lower layer) and on Si gel ( $C_{18}$ ) (MeOH-H<sub>2</sub>O; 3:2). A chromatographically homogeneous sample of **1** (10 mg) was obtained as a noncrystalline glass.

ACID HYDROLYSIS OF SAPONIN.—The saponin (1 mg) was refluxed with 5% methanolic HCl (5 ml) for 3 h. The neutralized product was partitioned between EtOAc and  $H_2O$ . The aglycone-containing organic phase was evaporated to dryness, silylated with BSTFA (0.1 ml) and pyridine (0.1 ml) at 50° for 20 min, and analyzed by gc.

SUGAR ANALYSIS.—The saponin (2 mg) was refluxed with HCOOH (90%, 1.0 ml) for 1 h at 100°, evaporated to dryness, and then refluxed in  $H_2SO_4$  (0.25 M, 1.5 ml) for 12 h at 100°. Deoxyhexose (200  $\mu$ l, 1 mg/ml) was added as internal standard, the mixture neutralized [Ba(OH)<sub>2</sub>, saturated solution], filtered, and evaporated to ca. 2 ml. The mixture was made alkaline (NH<sub>4</sub>OH, 2 M, 2 drops), then reduced with NaBH<sub>4</sub> (15 mg) at room temperature for 3 h. Dowex (H<sup>+</sup> form) ion exchange resin (5 ml) was added to neutralize the product, and, after filtration, the product was evaporated to dryness and acetylated with pyridine (0.1 ml) and Ac<sub>2</sub>O (0.1 ml) at 120° for 20 min and analyzed by gc.

PERMETHYLATIONS.—Compound 1 (6 mg) was dissolved in DMSO (0.5 ml) and treated with dimsyl sodium anion (2 M in DMSO, 0.5 ml) for 12 h under argon. MeI (0.5 ml) was added dropwise at 0°. After agitation in an ultrasonic bath for 1 h at room temperature, the mixture was added to H<sub>2</sub>O (5 ml), and any excess MeI was removed by passing a stream of argon through the solution. After extraction with CH<sub>2</sub>Cl<sub>2</sub> (4×3 ml) and washing with H<sub>2</sub>O (4×3 ml), the dried organic phase was evaporated to dryness and dissolved in C<sub>5</sub>D<sub>5</sub>N for <sup>1</sup>H nmr.

After <sup>1</sup>H nmr, the permethylated product was dissolved in  $CH_2Cl_2$ - $Et_2O$  (1:4, 2 ml) and reduced by refluxing the LiAlD<sub>4</sub> (15 mg) for 4 h. The cooled mixture was diluted with EtOAc-EtOH (1:1, 2 ml) and neutralized with H<sub>3</sub>PO<sub>4</sub> (2 M); it was then hydrolyzed, reduced, and acetylated as for sugar analysis (except that NaBD<sub>4</sub> was used in place of NaBH<sub>4</sub>) and the product analyzed by gc-ms.

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